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(71) Applicant: **CHIBA FLOUR MILLING CO. LTD.**
17, Shin-minato
Chiba City, Chiba (JP)
 Applicant: **Soma, Gen'ichiro**
1-10-21, Higashi-Tamagawa
Setagaya Ward, Tokyo (JP)
 Applicant: **Mizuno, Den'ichi**
18 Okamoto
Kamakura City, Kanagawa (JP)

(72) Inventor: **Soma, Gen'ichiro**
1-10-21 Higashi-Tamagawa, Setagaya Ward
Tokyo (JP)
 Inventor: **Yoshimura, Kiyoshi**
3-26-7 Isobe, Chiba City
Chiba (JP)
 Inventor: **Tsukioka, Daisuke**
1-21-17 Kasuga, Chiba City
Chiba (JP)
 Inventor: **Mizuno, Den'ichi**
Okamoto 18, Kamakura City
Kanagawa (JP)
 Inventor: **Oshima, Haruyuki**
2-10-513 Tate-machi 1097,
Hachioji City, Tokyo (JP)

(74) Representative: **Schrimpf, Robert et al**
Cabinet Regimbeau 26, Avenue Kléber
F-75116 Paris (FR)

(54) **Cholesterol-lowering agents and veterinary cholesterol-lowering agents.**

(57) A cholesterol-lowering composition comprising an effective amount of at least one member of LPS whose macrophage activation ED₅₀ is 0.4 - 100 ng/ml of culture solution in terms of its limulus test-positive LPS content observed on a sigmoid curve prepared by determining the ability of the LPS to activate the TNF productivity of macrophage cultured *in vitro*, and plotting the macrophage activation ability (%) along the axis of ordinate wherein the ability is estimated to be 0 % in the case where it corresponds to the quantity of TNF produced by macrophage with no LPS added thereto, and 100 % is assigned to the macrophage activation ability which provides the maximal and constant quantity of TNF produced by the macrophage and plotting the limulus test-positive LPS content of the LPS along the axis of abscissa on a logarithmic scale, in admixture with a pharmaceutically or veterinarily acceptable carrier, such that when administered to an animal, high cholesterol level of said animal is prevented or cured; and

a method of treating high cholesterol level of an animal comprising administration to said animal an amount of the above composition effective to prevent or cure the high cholesterol level of said animal.

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The present invention relates to cholesterol-lowering agents and veterinary cholesterol-lowering agents.

DESCRIPTION OF THE PRIOR ART

Cholesterol is a representative sterol found in vertebrates. It is found in almost all the cells of human beings, and is present in a free form or an ester with a fatty acid. From an epidemiological viewpoint, there is the correlation between serum cholesterol level and the frequency of occurrence of ischemic heart diseases. Clinical data shows that a decrease of 1 % of serum cholesterol level leads to a decrease of 2 % of the frequency of occurrence of coronary artery diseases. This decrease is reported to be more significant in more risky coronary artery diseases; see *"The newest therapy for circulatory diseases '86 to '87"* edited by Yasuda, et al., pp. 424 - 425, 1986, Nankohdo in Japan.

Therefore, various cholesterol-lowering agents have been developed. But, only nicotinic acid is evaluated to be effective against circulatory diseases; see *"The newest therapy for circulatory diseases '86 to '87" supra*, p. 425.

Nicotinic acid, however, has adverse effects, and further cannot be administered to patients suffering from severe hypotension or bleeding through the artery; see *"Commentary on the 11th Japanese Pharmacopeia"* supervised by Japanese Official Compendium Association, p. C-1207, 1986, Hirokawa-shoten in Japan.

BRIEF SUMMARY OF THE INVENTION

The present invention is intended to provide cholesterol-lowering agents and veterinary cholesterol-lowering agents which have a high degree of and lasting cholesterol-lowering effect but few adverse effects, and thus have a large therapeutic range, and further may be produced at a low cost, and may be administered orally, percutaneously or intravenously, and may be supplied in a large amount.

The cholesterol-lowering agents and veterinary cholesterol-lowering agents according to the present invention comprise at least one member of LPS whose macrophage activation ED_{50} is 0.4 - 100 ng/ml of culture solution in terms of its limulus test-positive LPS content observed on a sigmoid curve prepared by determining the ability of the LPS to activate the TNF productivity of macrophage cultured *in vitro*, and plotting the macrophage activation ability (%) along the axis of ordinate wherein the ability is estimated to be 0 % in the case where it corresponds to the quantity of TNF produced by macrophage with no LPS added thereto, and 100 % is assigned to the macrophage activation ability which provides the maximal and constant quantity of TNF produced by the macrophage (the quantity is abbreviated only as "maximal constant" in the other parts of the specification), and plotting the limulus test-positive LPS content of the LPS along the axis of abscissa on a logarithmic scale, if necessary or desired, in admixture with a pharmaceutically or veterinarily acceptable carrier.

An additional object of the present invention is to provide a method of treating high cholesterol level of an animal including human being, comprising administration to said animal an amount of at least one member of the LPS of the present invention effective to prevent or lower high cholesterol level, if necessary or desired, in admixture with a pharmaceutically or veterinarily acceptable carrier.

The term "therapeutic range" is the ratio of the maximum tolerant dose of the host to the medicine to the minimum effective dose of the medicine; the larger the ratio is, the better the medicine is.

"At least one member" is intended to mean that the respective members of the LPS of the present invention may be used individually or may be combined with each other or with any other matter optionally so far as their intended purpose is not missed.

"Macrophage" is the generic name for large amoeba-like cells which belong to immunocompetent cells, are present in most internal tissues of animals, and prey and digest particulate foreign matter and waste cells in the body.

"TNF" is the generic name for tumor necrosis factors produced by macrophage (*The Journal of Biol. Chem.*, 260, pp. 2345-2354, 1985), and the production quantity of TNF increases depending on the activity of macrophage.

"Limulus test" is a method invented by Levin in 1968 for quantitative determination of endotoxin using a horseshoe crab haemocyte extract and a chromogenic substrate.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 shows the behavior of wheat LPS and E. coli LPS available for use according to the present invention on SDS electrophoresis in comparison with those of standard markers.

Fig. 2 is a gas chromatographic chart of wheat LPS, showing the peaks evidencing the presence of fatty

acids therein.

Fig. 3 is a gas chromatographic chart of *E. coli* LPS, showing the peaks evidencing the presence of fatty acids therein.

Fig. 4 is a gas chromatographic chart of *B. pertussis* LPS available for use according to the present invention, showing the peaks evidencing the presence of fatty acids therein.

Figs. 5 to 8 are graphs evidencing the correlation between the macrophage activation ability and the limulus test-positive LPS content within the purview of the present invention.

Fig. 9 is a graph showing a cholesterol-lowering effect of the LPS according to the present invention.

DETAILED DESCRIPTION OF THE INVENTION

The LPS available for use as an active ingredient of a cholesterol-lowering agent or a veterinary cholesterol-lowering agent according to the present invention may be chosen regardless of its source, process for its production and purification or the like. For example, the LPS may be any one extracted from bacteria or plants, or may be a synthetic one such as synthetic lipid A. Here, throughout the specification and particularly in the claims, the LPS qualified with its source should not be interpreted to be restricted only those obtained from the specified source. Instead, it should be interpreted that the LPS include, in addition to those from the specified source, all the LPS obtained from bacteria or the like which grow on, in or together with the specified source during the course of its growth, storage, distribution or the like. For example, wheat LPS should be interpreted to include not only LPS from wheat, but also all the LPS obtained from bacteria or the like which grow on, in or together with wheat during the course of its growth, storage, distribution or the like. This is because it is well known in the art that, besides the hitherto known parasitic plants and animals, many types of organisms are allowed to grow on, in or together with certain plants, animals, or organisms belonging to mycota or lichenes.

Of the above-mentioned LPS, the LPS available for use as an active ingredient of a cholesterol-lowering agent or a veterinary cholesterol-lowering agent according to the present invention include those whose macrophage activation ED_{50} is 0.4 - 100 ng/ml of culture solution in terms of its limulus test-positive LPS content observed on a sigmoid curve prepared by determining the ability of the LPS to activate the TNF productivity of macrophage cultured *in vitro*, and plotting the macrophage activation ability (%) along the axis of ordinate wherein the ability is estimated to be 0 % in the case where it corresponds to the quantity of TNF produced by macrophage with no LPS added thereto, and 100 % is assigned to the macrophage activation ability which provides the maximal and constant quantity of TNF produced by the macrophage, and plotting the limulus test-positive LPS content of the LPS along the axis of abscissa on a logarithmic scale.

Limulus test-positive bacterial LPS

The hitherto known *E. coli* LPS, *B. pertussis* LPS, lipid A, etc. come under the captioned LPS.

E. coli LPS is commercially available from, for example, Difco Co. in U.S.A.

B. pertussis LPS is commercially available from, for example, Funakoshi Yakuhin (Funakoshi Pharmaceuticals Co.) in Japan. Alternatively, *B. pertussis* LPS may be prepared from dead cells of any publicly known *B. pertussis*, for example, Tohama I phase strain, by any of the publicly known processes described, for example, in the following literature:

Webster, et al., "*J. Immunol.*", 744, 55, 1955; and

Westphal, et al., "*Z. Naturforsch.*", 76, 148, 1952.

Lipid A is commercially available from, for example, Dai-ichi Kagakuyakuhin (Dai-ichi Chemicals Co.) in Japan.

Limulus test-positive vegetable LPS

The plants available for use as the starting material to prepare the cholesterol-lowering agents and veterinary cholesterol-lowering agents according to the present invention will be exemplified below.

For example, any plant belonging to Gymnospermae, Monocotyledoneae, Dicotyledoneae, Pteridophyta, algae or Fungi may be used separately or in admixture with each other.

The plants belonging to Gymnospermae available for use in the present invention include, for example, pine (*Pinus spp.*) belonging to Pinaceae.

Illustrative embodiments of the plants belonging to Monocotyledoneae available for use in the present invention are rice (*Oryza sativa* L.), wheat (*Triticum sativum* Lamarck, *Triticum aestivum* L.), barley, rye, oats (*Avena fatua*.), *Sasa albomarginata*, and pearl barley (*Coix Lacryma-jobi* L. var. *Ma-yuen* Stapf) belonging to Gramineae; blue flag (*Iris sanguinea* Donn., *Iris Nertschinskia* Lodd.) belonging to Iridaceae; garlic (*Allium sativum*)

vum L.), asparagus (*Asparagus officinalis* L.) and dwarf lilyturf (*Ophiopogon japonicus* Ker-Gawl. var. *genuinus* Maxim., *O. japonicus* Ker-Gawl.) belonging to Liliaceae; ginger (*Zingiber officinale* Roscoe), Japanese ginger (*Zingiber mioga*) and turmeric (*Curcuma domestica* Vahl, *Curcuma longa* L.) belonging to Zingiberaceae; jack-in-the-culprit (*Arisaema triphyllum*), etc.

5 The plants belonging to Dicotyledoneae available for use in the present invention are, for example, those belonging to Rubiaceae, Cruciferae, Cucurbitaceae, Lauraceae, Juglandaceae, Piperaceae, Umbelliferae, Menispermaceae, Saururaceae, Solanaceae, Rosaceae, Actinidiaceae, Leguminosae, Rutaceae, Magnoliaceae or Myristicaceae. More particularly, soybean (*Glycine Max* Merrill), "adzuki" bean (*Azuki angularis* Ohwi), broad bean (*Vicia* L.), kudzu (*Pueraria thunbergiana*), and licorice (*Glycyrrhiza glabra* L. var. *glandulifera* Regel et Herder) belonging to Leguminosae; potato (*Solanum tuberosum* L.), tomato (*Solanum lycopersicum* L., *Lycopersicon esculentum* Mill.), and red pepper (*Capsicum annuum* L.) belonging to Solanaceae; loquat (*Eriobotrya japonica* Lindl.) and peach (*Prunus persica* Batsch., *Prunus persica* Batsch. var. *vurgalis* Maxim.) belonging to Rosaceae; avocado (*Persea americana* Mill.) belonging to Lauraceae; walnut belonging to Juglandaceae; pumpkin (*Cucurbita moschata* Duch.) and *Gynostemma pentaphyllum* (Thunb.) belonging to Cucurbitaceae; *kaiware daikon* (Japanese radish) belonging to Cruciferae; silvewine (*Actinidia polygama* Maxim.) belonging to actinidiaceae; *Houttuynia cordata* Thunb. belonging to Saururaceae; pepper (*Piper nigrum* L.) belonging to Peperaceae; *Illicium verum* Hook. fil.; nutmeg (*Myristica fragrans* Houtt.) belonging to Myristicaceae; sour orange (*Citrus aurantium* L. subsp. *amara* Engl., *Citrus bigaradia* Risso et Poiteau) belonging to Rutaceae; "otane ninjin" (otane carrot) belonging to Araliaceae; *Seseli libanotis* Koch var. *daucifolia* DC. belonging to Umbelliferae; *Sinomenium acutum* Rehd. et Wils. belonging to Menispermaceae; *Uncaria rhychophylla* Miq., *Ouoruparia rhynchophylla* Matsum. belonging to Rubiaceae, etc. may be used.

The plants belonging to Pteridophyta available for use according to the present invention are, for example, horse tail (*Equisetum arvense* L.) belonging to Equisetaceae; royal fern (*Osmunda japonica* Thunb.) belonging to Osmundaceae, etc.

25 As plants belonging to Algae, any one belonging to, for example, Phaeophyceae, Rhodophyceae, Chlorophyceae or Cyanophyceae may be used separately or in admixture with each other. Illustrative embodiments of those belonging to Phaeophyceae are *Undaria pinnatifida* Suringar and kelp (*Laminaria japonica*) belonging to Laminariaceae; *Hizikia fusiformis* belonging to Sargassaceae, etc. Illustrative embodiments of those belonging to Rhodophyceae are asakusa laver (*Porphyra tenera*) belonging to Bangiaceae, etc. Illustrative

30 tive embodiments of those belonging to Chlorophyceae are chlorella (Chlorella) belonging to Oocystaceae, etc. As a fungus available for use according to the present invention, for example, any one belonging to Basidiomycetes or Ascomycetes may be used separately or in admixture with each other. Illustrative embodiments of those belonging to Basidiomycetes are *Lentinus edodes* Sing., *Cortinellus shiitake* P. Henn.; winter mushroom (*Flammulina velutipes*); *Lyophyllum shimeji*; *maitake* (phonetically spelled); *awabitake* (phonetically spelled) belonging to Polyporaceae; mushroom (*Agaricus bisporus*, *Agaricus bitorquis*); Jew's ear (*Hirneola auricula-judae* Berk., *Auricularia auricula-judae* Quel) belonging to auriculariaceae; *Pholiota nameko*. Illustrative embodiments of those belonging to Ascomycetes are baker's yeast and brewer's yeast belonging to Saccharomycetaceae. Here the "brewer's yeast" include those for brewing beer, sake (a Japanese alcoholic beverage) or wine or making *shoyu* (a Japanese soy sauce) or *miso* (a Japanese food paste made of soybeans, etc.), and further many other types of yeast belonging to Saccharomycetaceae and used for the preparation of whiskey, samshu (a Chinese alcoholic beverage) or the like. also *Cordyceps sinensis* Sacc., *Cordyceps sobolifera* belonging to Clavicipitaceae may be used according to the present invention.

45 Detection and quantitative determination of limulus test-positive LPS

The detection of the limulus test-positive LPS contained in any one of the plants referred to above and the determination of their LPS content may be carried out by using, for example, a reagent set commercially available from Sei-Kagaku Kogyo Co. in Japan under the trade name of Toxi Color System. That is, the starting plant is contacted with LS-1 set of said system, and the chromogenic strength of the plant is determined in comparison with the data of the calibration curve prepared using the Et-2 set of said system.

50 The vegetable LPS may be separated and purified in such a manner as mentioned below.

Separation and purification of limulus test-positive vegetable LPS

55 1) The starting plant is, if necessary after being sliced, dried and pulverized appropriately, suspended in distilled water, and then the supernatant is collected. For example, in case the starting plant is supplied in the form of cereal seeds, then the seeds are, if necessary or desired, after the removal of the seed coats, somewhat crushed or pulverized to an edible particle

size of powders. The resulting powders are prepared as a dispersion by addition of water thereto, and stirred. The dispersion is allowed to stand or subjected to centrifugation to remove the sediment, or the dispersion is worked into a dough by kneading, and is gently washed with water in a mixer to remove the sediment.

In the case where chlorella is used as the starting plant, then it is recommended first to crush the cell membranes, and then to wash the cells with ethanol to remove off the fat-soluble matter prior to the extraction with water.

When the above extraction is effected, there is no need to put limitations on the particle size of the starting plant, the temperature, the properties and the quantity of the water, the speed and the time of the stirring and the centrifugation conditions. These conditions may be adjusted conventionally depending on the starting plant used. Surely the water for extraction at a higher temperature tends to provide a larger quantity of a higher purity. For convenience only, however, the temperature of the water for the extraction is desired to be not more than 50°C so that the starch in the starting plant is not gelatinized. In addition, though the quantity of the water to be added changes depending on the type and the particle size of the starting plant used, for easiness of operation only, it is desired to be 20-50 w/v %. At the end of this stage of operation, the purity of the limulus test-positive vegetable LPS plant of the present invention increases to about 30-fold value in the case of wheat seeds judging from the data on limulus test activity.

Hereunder, the description of the specification will be made with particular reference to the case where cereal seeds are used as the starting material. But, no doubt it is sure that the teachings found in such description are sufficient for any person skilled in the art to remove off sugars, proteins and other useless components from another starting material thereby extracting the limulus test-positive LPS of a high purity available for use according to the present invention.

2) In order to get a higher purity, the supernatant from the above 1) may be subjected to ultrafiltration in the conventional manner to remove fractions having molecular weights of 5,000 or less.

3) The resulting dried sample is then suspended in distilled water to a proportion of 50 mg/ml after which it is subjected to centrifugation to collect the supernatant.

4) The supernatant from the above 3) is cooled on ice water, and then an acid added thereto to produce sediment. Here, any acid may be employed; for example, trichloroacetic acid (hereunder referred to as TCA only), perchloric acid, trifluoroacetic acid, acetic acid or dichloroacetic acid may be employed.

5) Then the mixture is subjected to centrifugation to collect the sediment which is then washed with distilled water, and then is subjected to centrifugation again to collect the sediment.

6) The sediment from the above 5) is suspended in distilled water, and an alkali is added to the suspension until the sediment dissolves therein. Here, any alkali may be used; for example; sodium hydroxide, potassium hydroxide, ammonia, sodium carbonate, sodium acetate or the like may be used. Here, care should be taken that the basicity of the suspension does not go over pH 11 when the sediment dissolves in order to keep the object LPS from being deactivated.

7) Then an acid is added to the suspension to bring its pH to pH 8 followed by warming to 37°C. Further addition of an acid makes the suspension basic to produce sediment which is then collected by a centrifuge warmed at 37°C.

8) The thus collected supernatant is cooled on ice, and then subjected to centrifugation at 4°C again.

9) The supernatant is collected, and then neutralized by the addition of an alkali, and concentrated by ultrafiltration in the conventional manner. Here, any alkali may be used.

10) Then the concentrate is subjected to gel filtration in the conventional manner, and limulus test-positive fractions are collected and combined. Here, the support for the gel filtration may be, for example, Sephadex G-75, G-100, Sephacryl S-200, Sepharose 6B (the foregoing are all manufactured by Pharmacia Inc. in U.S.), Biogel P-100 (manufactured by Biorad Inc.), Toyo Pearl HW-50, HW-55 (manufactured by Toyo Soda Kogyo Co. in Japan), or the like. The buffer solution may be any one as long as it can keep the pH within the range of 3-10. For example, Tris-HCl or phosphate buffer solutions may be used.

11) Then a proteolytic enzyme is added to the combined fractions which are then incubated at 37°C for 2 or more hours to decompose the remaining proteins. The thus treated solution is then concentrated by ultrafiltration in the conventional manner. Here, no specific proteolytic enzyme is required, and, for example, V8 protease, chymotrypsin, trypsin or thermolysin may be used separately or in optional combinations thereof. The commercially available proteolytic enzymes which may be used according to the present invention include, for example, Pronase[®] (Kaken Kagaku Co. in Japan) and Proteinase[®] (Merc Co. in U.S.).

12) Then the collected fractions are treated conventionally, for example, by being subjected to ion-exchange chromatography using mono Q Sepharose or Q Sepharose manufactured by Pharmacia Inc., to collect the limulus test-positive fractions.

13) Then, the fractions are subjected to gel filtration conventionally to collect the desalted limulus test-positive fractions.

By the above-mentioned procedures, in the case of wheat seeds, about 20% of the originally present limulus test-positive matter is recovered, and a purified sample of about 95% purity is obtained. This value achieved in the case of wheat seeds is about 1,000 times as high as that obtained at the end of 1) referred to above.

The limulus test-positive vegetable LPS prepared according to the above procedures and other LPS within the purview of the present invention may be supplied as such, or in the forms diluted to an optional desired degree. In addition, in order to improve its stability, they may be supplied as dried powders in the conventional manner including lyophilization and spray drying. Each of these forms may be produced conventionally.

Determination of ability of the LPS according to the present invention to activate *in vitro* TNF productivity of macrophage

15 Carswell et al. report that priming and triggering steps are necessary to produce endogenous TNF in the body of an animal; see *Proc. Natl. Acad. Sci. USA.*, 72, pp. 3666 - 3670, 1975. Thereafter, many candidate chemicals were tried to stimulate the respective steps. The chemical used to start the priming step is a primer (endogenous TNF production stimulator), while that administered to start the triggering step is a trigger (endogenous TNF productive agent).

20 In order to determine the ability of LPS to activate the *in vitro* TNF productivity of macrophage, a recombinant mouse IFN- γ is added to peritoneal macrophage cells of mouse as the primer followed by addition of the LPS as the trigger thereto, and then the TNF activity is determined.

The TNF activity is determined, as follows, on the basis of the cytotoxicity to L929 cells (*Proci. Natl. Acad. Sci. U.S.A.*, 72, pp. 3666 - 3670, 1983). L-929 cells are cultured in Eagles' Minimum Essential Medium (hereunder referred to only as MEM) with 5 % fetal calf serum (hereunder referred to only as FCS) added thereto until 100 μ l of the medium contains 8×10^4 cells, and then the cells are grown in a flat-bottomed plate having 96 wells.

25 The growth conditions are 37°C in the presence of 5 % CO₂, and under a humidity of 100 % for 2 hours, and the procedures may be the same as for the conventional cell culture. Then actinomycin D is added to the medium to a final concentration of 1 μ g/ml, and the volume of the culture solution is adjusted to 150 μ l. Immediately thereafter 50 μ l of the sample diluted appropriately with MEM medium is added to the culture solution. Here, ED₅₀ may be determined by adjusting the dilution appropriately. The L-929 cells having a final volume of 200 μ l are cultured for an additional 18 hours under the same conditions as described above.

35 In order to determine the cell necrosis activity, first the whole medium is removed followed by addition of a 1% methyl alcoholic solution containing 0.1 % crystal violet for fixation staining. Crystal violet stains all the eukaryotic cells, but the dead cells are removed off from the bottom of the flask only by washing after the staining; so the cell necrosis activity may be determined directly. The staining degree is measured on the basis of adsorption at OD_{590nm}, and is compared with that of a control to determine the cell necrosis activity. This activity is defined as follows.

40 The dilution of the sample which allows 50 % of the L-929 cells to survive (N) is determined. Rabbit TNS is used as the control, and its activity n (units/ml) is determined using 2.4×10^6 units/mg/ml of TNF- α . The dilution which provides ED₅₀ of rabbit TNS is determined.

The activity of the sample (units/ml) is calculated by the equation $N/C \times n$.

45 Forms supplied

The cholesterol-lowering agents and veterinary cholesterol-lowering agents according to the present invention may be supplied conventionally in the form of powders, granules, pills, tablets, troches, capsules, solutions, pastes, ointments, liniments, lotions, suppositories, injections, etc. For veterinary use, also the agents may be prepared in the form of feed additives, premix preparations, drinking water additives. Here, the "premix preparations" are such preparations as contain feed components beforehand so that they are easily mixed in the feed. The feed additives are preferred to be powders or granules. Any commercially available feed may be used to prepare the above-mentioned feed additives, premix preparations, etc. The feed may contain minerals, vitamins, amino acids and any other feed additives.

55 If desired, these preparations may contain excipients, preservatives, buffers, etc. conventionally to improve the shelf life, homogeneity, etc. In addition, the preparations may contain correctives to improve taste, odor, appearance, etc. The excipients include, for example, lactose, starch, etc. The preservatives include, for example, parahydroxybenzoic esters such as methyl, ethyl or propyl paraoxybenzoate, sodium dehydroace-

tate, phenols, methyl, ethyl or propylparabene, etc. The buffers include, for example, citric, acetic or phosphoric acid salts, etc.

Hereunder, the present invention will be explained in detail with reference to reference examples, examples and experiments.

Reference Example 1 (preparation of wheat LPS)

1) In a small kneader, there was charged 3,120 g of hard flour containing 1.09% of ash (Hard Red Spring wheat from U.S.A. or Canada) followed by addition of 2.03ℓ of distilled water thereto and kneading for 10 minutes to prepare a dough. The mixture was allowed to stand for 15 minutes, and then 10ℓ of water was added to the mixture followed by gentle stirring to extract a starch emulsion and to dissolve soluble ingredients in the water. The resulting solution was allowed to stand in a refrigerator at 5°C for 12 hours, and then the sediment containing starch etc. was removed. The supernatant was lyophilized to get 201.1 g of powders (Powder A).

Then 5 ℓ of distilled water was added to the residual dough followed by gentle stirring, and the mixture was treated in the same manner as the above to get 40.1 g of powders (Powder B).

2) The powders A and B were charged in an ultrafilter HF-Lab1 (Amicon Co.), and the ultrafiltration was carried out using a hollow cartridge HF-Lab1PM5 for fractions having a molecular weight of 5,000, and another hollow cartridge HF-Lab1PM10 for fractions having a molecular weight of 10,000; the temperature range was 5 - 10°C, the inlet pressure was 25 p.s.i. (1.76 kg/cm²), and the outlet pressure was 15 p.s.i. (1.06 kg/cm²).

As a result, the respective fractions were named as follows:

- Powder A: Fractions having a molecular weight of 5,000 or less were named a₁.
Fractions having a molecular weight of 5,000 or more were named a₂.
- Powder B: Fractions having a molecular weight of 5,000 or less were named b₁.
Fractions having a molecular weight of 5,000 or more were named b₂.
- Powder A: Fractions having a molecular weight of 10,000 or less were named a₃.
Fractions having a molecular weight of 10,000 or more were named a₄.
- Powder B: Fractions having a molecular weight of 10,000 or less were named b₃.
Fractions having a molecular weight of 10,000 or more were named b₄.

Each of these fractions was subjected to the limulus test according to the method detailed in Experiment 1 given later, and it was confirmed that much limulus test-positive ingredients are present in fractions having a molecular weight of 5,000 or more, whereas little limulus test-positive ingredients are present in fractions having a molecular weight of 5,000 or less.

3) Thirty grams of the above powder a₂ was placed in a 1 ℓ Erlenmeyer flask after which 600 ml of distilled water was poured thereon. The resulting mixture was stirred with a stirrer for 60 minutes, and then subjected to centrifugation at 10,000 G for 10 minutes using a Hitachi cooling type high speed centrifuge SCR-20B (the rotor RPR16 was cooled to 4°C beforehand) to recover the supernatant.

4) The supernatant from the above 3) was placed in another 1 ℓ Erlenmeyer flask followed by dropwise addition of 20.5ml of a 100 % aqueous solution of TCA cooled to 2°C beforehand while cooling on ice (the temperature of the solution was about 2°C) and stirring with a stirrer. After the completion of the dropwise addition, the flask was allowed to stand in ice water for 10 minutes.

5) Next, the mixture was subjected to centrifugation (at 10,000 G for 10 minutes) at 4°C in the same manner as the above to collect sediment which was in turn put in a 500 ml beaker together with 300 ml of distilled water while cooling in ice water to prepare a suspension. The resulting suspension was cooled in ice water and subjected to centrifugation (at 10,000 G for 10 minutes) at 4°C in the same manner as the above to recover sediment.

6) The sediment was placed in a 1 ℓ beaker followed by addition of 500 ml of distilled water to prepare a suspension. The resulting suspension was neutralized to pH 7 by the use of about 3.5ml of 1 N sodium hydroxide, and then, while cooling in ice water, about 2 ml of 1 N sodium hydroxide was added to the neutralized suspension until a 0.02 N sodium hydroxide solution is prepared to dissolve the sediment.

7) About 1.5 ml of 1 N hydrochloric acid was added to the solution to bring the pH to 8 followed by addition of 100 ml of distilled water. The solution was transferred to a 1 ℓ Erlenmeyer flask which was then shaken slowly in an incubator at 37°C for 30 minutes.

8) The solution from the above 7) was mixed with 30 ml of a 100 % aqueous solution of TCA, and then slowly shaken in an incubator at 37°C for 10 minutes. Thereafter the solution was subjected to centrifugation at 3,000 G for 10 minutes using a centrifuge, Tomy CD100R (Tomy Seiki Co. in Japan)

9) The supernatant was collected and cooled on ice, and then subjected to centrifugation at 4°C (at 10,000 G for 10 minutes.)

10) The supernatant was collected and neutralized to pH 7 with about 3.6 ml of 10 N sodium hydroxide,

and the neutralized solution was concentrated using an ultrafilter (Toyo Roshi UHP-150, Filter:UK-10, N₂ pressure: 4.0 kg/cm²)

11) The resulting concentrate (60 ml) was subjected to gel filtration (buffer: 10 mM Tris-HCl / 10 mM NaCl (pH7.5), flow speed: 60 ml/h.) to collect 20 ml fractions using Sepharose 6B column (manufactured by Pharmacia Inc., column size: 5 cm (i.d.) × 100 cm (2l)).

12) The 43rd to 56th fractions were combined to prepare 280 ml of a solution followed by addition of 450 µg of Pronase E (Kaken Kagaku Co. in Japan) and warmed at 37°C for 2 hours while shaking. Thereafter the mixture was concentrated using an ultrafilter (Toyo Roshi UHP-62, filter: UK-10, N₂ pressure: 4.0 kg/cm²). Then the concentrate was subjected to anion exchange chromatography using FPLC system (manufactured by Pharmacia Inc., column: mono Q HR 10/10). That is, the sample was applied to the column using a buffer solution containing 10 mM Tris-HCl (pH7.5) and 10mM of NaCl, and then the column was washed with 200 ml of the same type buffer solution as the above but containing an increased amount 165 mM of NaCl. Then, totally 400 ml of the eluate was used to elute the object LPS while increasing the NaCl concentration so that the NaCl concentration gradient ranges from 165 mM to 1 M. Two ml fractions were collected. The 5th to 8th fractions after the start of the concentration gradient which were confirmed to be positive to the limulus test were combined to yield 8 ml of LPS of about 92 % purity (LPS: 3.03 mg (in terms of E. coli LPS according to the limulus test; the same applies to the following), sugar: 0.23 mg, protein: 0.04 mg)

13) Then, the 8 ml was subjected to gel filtration (buffer: water) using Sephadex G-25 (column: 2.0 cm (i.d.) × 20.2 cm (66 ml)) to collect 3 ml fractions. The 9th to 12th fractions confirmed to be positive to the limulus test were combined to yield 12 ml of glycolipid of about 95 % purity (LPS: 2.7 mg, sugar: 0.18 mg, protein: 0.03 mg). The sugar content was determined according to the phenol - sulfuric acid method, while the protein content, according to the Lowry method. Here, the combined fractions were confirmed to be acidic by anion exchange chromatography. The molecular weight according to SDS electrophoresis was 6,000 - 10,000.

14) The above fractions were frozen at -80°C, and then lyophilized to a constant weight which was measured to be 0.75 mg. Hereunder, this lyophilized sample is referred to as wheat LPS only.

The limulus activity of wheat LPS was determined to be 2.7 mg according to the method described in Experiment 1 given later, so its specific activity is calculated to be 3.6 (2.7 ÷ 0.75). Here, the foregoing purification procedures are supposed to have removed substantially all the independent sugars present as contaminants, so all the sugars detected are supposed to be those constituting the wheat LPS. Thus, the purity of the wheat LPS on the basis of weight at this stage may be calculated as follows:

Protein content: 0.03 mg

LPS content: 0.72 mg (0.75 - 0.03)

Purity = $0.72 \div 0.75 \times 100 = 96 (\%)$

Physical properties of wheat LPS

15) Molecular weight

Wheat LPS was dissolved in distilled water to prepare a 1 mg/ml solution of CHF, and 4 µl of the solution was placed in a 1.5 ml Treff tube. Separately, 2.5 % of SDS, 5 % of mercaptoethanol and 10 mM Tris - HCl (pH8.0) were added to 1 mM EDTA to prepare 1 µl of an SDS treatment solution, and this solution was added to the above solution in the Treff tube which was then dipped in boiling water for 3 minutes. Phast System of Pharmacia Inc. was used in the electrophoresis experiments. That is, one µl of the mixture was applied to a gel (Phast Gel Gradient 8-25 of Pharmacia Inc.) which was connected to the electrodes through SDS-Buffer Strip of Pharmacia Inc., the maximum potential difference and the maximum electric current were set to 250 v and 10 mA, respectively, and then the electrophoresis started.

At the end of the electrophoresis, the behaviors in Coomassie staining and silver staining were observed.

For Coomassie staining, Phast Gel Blue R was used as the staining solution, and a mixture of methanol, acetic acid and distilled water (volumetric ratio is 3:1:6), as the decoloring solution. The staining and decoloring were carried out in the following order:

1) Stained at 50°C for 8 minutes.

2) Stained at 50°C for 5 minutes.

3) Stained at 50°C for 8 minutes.

4) Decolored at 50°C for 10 minutes.

5) Protected at 50°C for 5 minutes (a mixture of glycerol, acetic acid and distilled water in a volumetric ratio of 5:10:85).

6) Dried.

The silver staining was carried out in the following order:

- 1) Treated with a wash (a mixture of ethanol, acetic acid and distilled water in a volumetric ratio of 5:1:4) at 50°C for 2 minutes.
- 2) Treated with a wash (a mixture of ethanol, acetic acid and distilled water in a volumetric ratio of 10:5:85) at 50°C for 2 minutes.
- 5 3) Treated with a wash (a mixture of ethanol, acetic acid and distilled water in a volumetric ratio of 10:5:85) at 50°C for 4 minutes.
- 4) Treated with a sensitizer solution (8.3 % glutar dialdehyde) at 50°C for 6 minutes.
- 5) Treated with a wash (a mixture of ethanol, acetic acid and distilled water in a volumetric ratio of 10:5:85) at 50°C for 3 minutes.
- 10 6) Treated with a wash (a mixture of ethanol, acetic acid and distilled water in a volumetric ratio of 10:5:85) at 50°C for 5 minutes.
- 7) Treated with a wash (deionized water) at 50°C for 2 minutes.
- 8) Treated with a wash (deionized water) at 50°C for 2 minutes.
- 9) Treated with 0.25 w/v % of silver nitrate at 40°C for 13 minutes.
- 15 10) Treated with a wash (deionized water) at 30°C for 30 seconds.
- 11) Treated with a wash (deionized water) at 30°C for 30 seconds.
- 12) Treated with a developer (0.04 v/v % of formaldehyde + 2.5 w/v % of sodium carbonate as a wash) at 30°C for 30 seconds.
- 13) Treated with a developer (0.04 v/v % of formaldehyde + 2.5 w/v % of sodium carbonate as a wash) at 30°C for 4 minutes.
- 20 14) Treated with a reaction termination solution (5 v/v % of acetic acid) at 50°C for 2 minutes.
- 15) Treated with a protective solution (a mixture of acetic acid, glycerol and distilled water in a volumetric ratio of 10:8:85) at 50°C for 3 minutes.
- 16) Dried.

25 LPS are subjected to silver staining, but not to Coomassie staining. This difference was taken into consideration to observe the stained bands, and the main stained band of wheat LPS was found at a position indicating a molecular weight of $8,000 \pm 1,000$. Similarly, the stained bands of *E. coli* LPS (0128:B8 commercially available from Difco Co. in U.S.A.) were observed to be successive in tiers, and the molecular weight of the stained band having the highest staining strength was estimated to be $30,000 \pm 5,000$.

30 The stained bands of wheat LPS and *E. coli* LPS observed on SDS electrophoresis are shown in Fig. 1 in comparison with those of standard markers 1-6 corresponding to molecular weights of 14,400, 20,100, 30,000, 43,000, 67,000 and 94,000, respectively.

35 16) Phosphorus content

The captioned content was determined as follows according to the Chen-Toribara method (Chen et al., "Analytical Chemistry", vol. 28, pp. 1756 - 1758, 1956)

40 Wheat LPS was dissolved in distilled water to prepare 20 μ l of a solution containing 25 μ g of wheat LPS which was then placed in a small test tube. To the mixture there was added 20 μ l of 50 v/v % perchloric acid, and then the mixture was heated on a gas burner for 1 minute to ash. Thereafter, 0.5 ml of distilled water and then 0.5 ml of a reaction reagent (a portion of the preparation made by mixing 1ml of 6N sulfuric acid, 2ml of distilled water, 2 ml of 2.5 v/w % ammonium molybdate and 1ml of 10 v/w % of ascorbic acid) were added to the heated mixture which was then allowed to stand for 30 minutes at room temperature. Thereafter the absorption at 820nm (OD_{820nm}) was determined. Here, as the standard sample for the preparation of the calibration

45 curve, potassium dihydrogen phosphate (manufactured by Wako Jun-yaku Co. in Japan) was diluted with water to prepare 0.5 ml of solutions containing 2.5 μ g, 1 μ g, 0.25 μ g or 0 μ g of the standard in terms of phosphorus. In this connection, 1 g of phosphorus corresponds to 4.39 g of potassium dihydrogen phosphate. The effects observed are shown in Table 1 given below.

Table 1

O D _{520nm}	Samples
	Potassium dihydrogen phosphate (in terms of phosphorus; μ g)
0 . 0 0 2	0
0 . 1 5 0	0 . 2 5
0 . 6 2 0	1 . 0
1 . 5 5 9	2 . 5
	Wheat LPS (data of four samples) (content of phosphorus calculated considering calibration curve; μ g)
0 . 0 3 6	0 . 1
0 . 0 7 3	0 . 2
0 . 1 0 4	0 . 3
0 . 1 3 9	0 . 4

Note: The data of wheat LPS are modified by subtracting the values of the control not subjected to the heating from the observed values in order to avoid occurrence of errors due to mixing-in of inorganic phosphorus from, for example, phosphate buffer solution.

On the assumption that the molecular weight of wheat LPS is 8,000, the number of phosphorus in LPS is calculated to be 1 - 4 per mol. according to the following equation on the basis of the data shown in the above table .

$$\text{Phosphorus content} \times 10^{-6} \times \frac{\text{Molecular weight}}{25 \times 10^{-6}} \times \frac{1}{32}$$

For an explanation as to why the number of phosphorus ranged from 1 to 4, one may guess that the phosphoric acid (s) may be eliminated due to the mixing-in of the monophosphoesterase in the purification stage. In view of this, it may be right to conclude that wheat LPS has not less than 1 phosphorus per mol.

17) Hexosamine content

The captioned content was determined as follows according to the Elson-Morgan method (Library of biochemical experiments, No. 4, pp. 377 - 379, Tokyo Kagaku Dojin Shuppan Co. in Japan).

Wheat LPS was dissolved in distilled water to prepare a solution containing 1 mg/ml of wheat LPS, and its 100 μ l portion was placed in a test tube with a screwcap (manufactured by Iwaki Glass Co. in Japan) followed by addition of 100 μ l of 8N HCl thereto, and the mixture was heated at 110°C for 16 hours, and then

about 200 $\mu\ell$ of 4N NaOH was added to the mixture to bring the pH to 7. A 100 $\mu\ell$ portion of the mixture was separated off and placed in another test tube with a screwcap followed by addition of 200 $\mu\ell$ of Reagent A explained below thereto. The mixture was then heated at 105°C for 1.5 hours, and then cooled with a running water. Next, a 100 $\mu\ell$ portion of the mixture was separated off followed by addition of 670 $\mu\ell$ of a 96 % ethanol and then 67 $\mu\ell$ of Reagent B explained below, and was then allowed to stand at room temperature for 1 hour followed by determination of adsorption at 535nm. As the standard sample to prepare the calibration curve, 0.20 - 200 $\mu\text{g/ml}$ of N-acetyl glucosamine (Wako Jun-yaku Co. in Japan) was used. Reagent A: prepared by mixing 75 $\mu\ell$ of acetyl acetone and 2.5 ml of 1.25 N sodium carbonate

Reagent B: prepared by mixing 1.64 of p-dimethyl benzaldehyde, 30 ml of conc. hydrochloric acid and 30 ml of 96% ethanol

As a result, the number of the hexosamine in wheat LPS was 6 ± 2 per mol. on the assumption that its molecular weight is 8,000.

17) Fatty acid content

To 90 $\mu\ell$ of a solution of wheat LPS in distilled water containing 1 mg/ml of wheat LPS there was added 10 $\mu\ell$ of an internal standard (0.55 mM margaric acid) followed by addition of 1.0 ml of 0.5M sodium methylate for hydrolysis and esterification of fatty acid esters. The mixture was allowed to stand at room temperature for 1 hour followed by the addition of 960 $\mu\ell$ of 0.5 N HCl thereto for neutralization. Two ml of hexane was added to the mixture which was then stirred vigorously for 15 minutes. Next, the resulting mixture was subjected to centrifugation at 1,000 G for 5 minutes to separate off the hexane layer. The hexane was evaporated off by nitrogen gas, and the layer was concentrated to about 20 $\mu\ell$. The resulting concentrate was subjected to gas chromatography (GC8APF manufactured by Shimazu Co. in Japan; capillary column: FSCAP Sp2330 manufactured by Spelco Co. in Canada; carrier gas: nitrogen) to determine the fatty acid content. As the standard for determination of the fatty acid content, there was used E. coli type LA-15-PP, a synthetic lipid A manufactured by Dai-ichi Kagaku Yakuhin Co. in Japan and known to have a molecular weight of 2,000 and a fatty acid count of 6 per mol.

As a result, the number of the fatty acid in wheat LPS was 6 ± 2 per mol. on the assumption that its molecular weight is 8,000.

The charts observed in the above gas chromatography are shown in accompanying Figs. 2 - 4. Fig. 2 is the chart of wheat LPS, Fig. 3 is that of E. coli LPS, and Fig. 4 is that of B. pertussis LPS.

The retention times (minutes) corresponding to the main peaks shown in Figs. 2 - 4 are as follows:

	<u>No. of peak</u>	<u>Retention time (min.)</u>
Fig. 2:	1	2 . 4 5 0
	2	2 . 7 5 8
Fig. 3:	1	2 . 4 1 7
	2	2 . 7 4 2
Fig. 4:	1	2 . 4 3 3
	2	3 . 0 2 8

By the comparison of Figs. 2 - 4, it is apparent that wheat LPS and E. coli LPS show similar charts, whereas the chart of wheat LPS is clearly different from that of B. pertussis LPS.

18) KDO content

The KDO content was determined as follows on the basis of the diphenylamine method (Shaby R. et al., "Analytical Biochem.", 58(1), pp. 123-129, 1974).

A KDO detection reagent was prepared by combining 500 mg of diphenylamine, 5 ml of ethanol, 45 ml of glacial acetic acid and 50 ml of conc. hydrochloric acid (all commercially available from Wako-junyaku Co. in Japan). a 500 μ l portion of the prepared reagent was combined with 250 μ l of distilled water containing 1.05 mg/ml of wheat LPS. The resulting mixture was heated in a boiling water bath at 100°C for 30 minutes and then cooled in cooling water at 23°C for 30 minutes. The UV absorption of the mixture was determined at 420, 470, 630 and 650 nm to provide data A_{420} , A_{470} , A_{630} and A_{650} , respectively. As the standard sample, there was used 250 μ l of distilled water containing 127 μ g/ml of ammonium salt of KDO (Sigma Co. in U.S.A.). The value S for the test and standard samples was calculated according to the following equation:

$$S = A_{420} - A_{470} + A_{630} - A_{650}$$

The value of the test sample (S_t) was 0.379, whereas that of the standard sample (S_s) was 0.294. The comparison of the two values suggests that wheat LPS contains 5 ± 1 mol. on the assumption that its molecular weight is 8,000.

Reference Example 2 (preparation of chlorella LPS)

1) Thirty grams of cell membrane-crushed chlorella (Mannan-foods YS Co. in Japan) was washed with ethanol until the wash was not colored green any more.

2) The residue (26 g) was dissolved in distilled water to prepare a 100 mg/ml solution which was then shaken at 45°C for 2 hours followed by centrifugation (4°C, 10,000 G for 30 min.).

3) The supernatant was filtered through Toyo-roshi No. 2 followed by extraction with distilled water.

4) The extracts (290 ml) were subjected to anion exchange chromatography under the following conditions:

Column: Q Sepharose (\varnothing 3 cm x 23 cm, volume: about 180 ml)

Buffer: 10 mM Tris-HCl (pH 7.5)

Gradient of NaCl conc.: 10 mM, 400 mM, 1M

Flow rate: 100 - 200 ml/hr.

Temperature: Room temperature

5) The eluate (310 ml) was treated with glucoamylase at pH 5.0, 40°C, for about 2 hrs. to decompose the starch. The decomposition of the starch was confirmed by the fact that the coloring of the solution was not found in the starch iodide reaction.

6) The solution was subjected to centrifugation (10,000 G for 10 min.) to collect the supernatant which was then neutralized with a 10 N NaOH solution to pH 7 followed by ultrafiltration using a ultrafilter having a pore size which excludes matter having a molecular weight of 20,000 or more.

7) The thus prepared concentrate (30 ml) was subjected to anion exchange chromatography using FPLC system (column: mono Q HR 10/10). That is, the sample was applied to the column using a buffer solution containing 10 mM Tris-HCl (pH 7.5) and 10mM of NaCl, and then the column was washed with 200 ml of the same type buffer solution as the above but containing an increased amount 165 mM of NaCl. Then, totally 400 ml of the eluate was used to elute the object LPS while increasing the NaCl concentration so that the NaCl concentration gradient ranges from 165 mM to 1 M. Two ml fractions were collected. The 5th to 8th fractions after the start of the concentration gradient which were confirmed to be positive to the limulus test were combined.

8) Then, a 8 ml portion of the fractions was subjected to gel filtration (buffer: water) using Sephadex G-25 (column: 2.0 cm (i. d.) x 20.2 cm (66 ml)) to collect 3 ml fractions. The 9th to 12th fractions confirmed to be positive to the limulus test were combined to yield a 12 ml solution (LPS: 14.3 mg, sugar: 2.0 mg, protein: 0.53 mg). The LPS content was determined by the procedure as described in Experiment 1 mentioned later, the sugar content was determined according to the phenol - sulfuric acid method, and the protein content, according to the Lowry method.

9) The above fractions were frozen at -80°C, and then lyophilized to a constant weight which was measured to be 5.8 mg. Hereunder, this lyophilized sample is referred to as chlorella LPS.

The limulus activity of chlorella LPS corresponds to 14.3 mg, so its specific activity is calculated to be 2.5 (14.3 \div 5.8).

The foregoing purification procedures are supposed to have removed substantially all the independent sugars present as contaminants, so all the sugars detected are supposed to be those constituting chlorella LPS. Thus, the purity of chlorella LPS on the basis of weight at this stage may be calculated as follows:

Protein content: 0.53 mg

LPS content 5.27 mg (5.8 - 0.53)

purity = 91 (%) (5.27 \div 5.8 x 100)

Physical properties of chlorella LPS

Following the procedures as described in Reference Example 1, the captioned properties were determined as follows:

- 5 Molecular weight: 40,000 - 90,000
 Phosphorus content: 4 ± 1 per molecular weight of 10,000
 Hexosamine content: 7 ± 1 per molecular weight of 10,000
 Fatty acid content: 6 ± 1 per molecular weight of 10,000
 KDO content: 2 ± 1 per molecular weight of 10,000

Reference Example 3 (preparation of B. pertussis LPS)

An experimental B. pertussis solution obtained from Serum Laboratory, a public institute of Chiba prefecture in Japan (2.0×10^{10} cells / ml) was used.

- 15 The solution was suspended in sterile water to prepare a suspension containing 25 mg (dry basis) / ml of dead cells. To the suspension, there was added an equivalent of a 90% hot phenol solution (68 - 70°C) was added, and the mixture was shaken at 68°C for 1 hr. The mixture was subjected to centrifugation at 8,000 G, 4°C for 20 min. to collect the aqueous layer. Sterile water in the same quantity as of the aqueous layer was added to the remaining phenol, and the mixture was shaken in the same manner as the above. The resulting
 20 aqueous layer was combined with the first aqueous layer followed by dialysis in running water overnight, and then the mixture was concentrated to a tenth using a rotary evaporator. The concentrate was subjected to centrifugation at 8,000 G, 4°C for 20 min. The supernatant was separated off, and a small amount of sodium acetate was added thereto. Cold ethanol at 0 - 4°C was added to the mixture in an amount of six times as much as the latter, and the resulting mixture was allowed to stand at -20°C overnight. Then the mixture was subjected to
 25 centrifugation at 4,000 G, 4°C for 30 min. to collect the sediment which was subjected to centrifugal washing with ethanol (twice) and acetone (once) followed by drying with an aspirator. The residue was suspended in distilled water to prepare a 20 mg / ml of solution which was then subjected to ultrasonic treatment with a Sonifia 185 (Branson Co. in U.S.A.) (outlet control 5, 15 min., room temperature). The solution was subjected to centrifugation at 2,500 G, 4°C for 10 min. to separate off the supernatant.

- 30 The supernatant was treated at 4°C with nucleases, DNase I and Rnase A (both manufactured by Sigma Co. in U.S.A) for 15 - 16 hrs; totally 10 µg/ml of DNase I and 20 µg/ml of Rnase a were used. The same amount of the nucleases as the above were added to the mixture followed by warming at 37°C for 2 hrs and centrifugation at 2,500 G, 4°C for 10 min. to separate off the supernatant.

- The supernatant was filtered through a pore size of 0.2 µm using Acrodisc manufactured by Gelman Co. in U.S.A. The filtrate was subjected to molecular sieve (resin: Sepharose 6B manufactured by Pharmacia Co. in U.S.A; column size: 5 cm (i.d.) x 100 cm (length); buffer: 10 mM of Tris-HCl / 10 mM of NaCl (pH 7.5); flow rate: about 3 ml/cm²/hr.). The fractions confirmed to be positive to limulus test with LS-1 kit commercially available from Sei-Kagaku Kogyo Co. in Japan were collected and filtered through a pore size of 0.2 µm using Acrodisc mentioned above. The filtrate was subjected to ion exchange (apparatus: FPLC manufactured by Pharmacia
 40 in U.S.A.; resin: mono Q HR 10/10 manufactured by Pharmacia in U.S.A.; buffer: 10 mM of Tris-HCl / 10 mM of NaCl (pH 7.5); flow rate: 2 ml/min.) wherein the filtrate was washed with the buffer for 15 min., then, after the NaCl content of the buffer was increased to 165 mM, for 30 min., then, for 20 min. while increasing the NaCl content to provide a NaCl content gradient from 165 mM to 1 M, and then, for 30 min. at the NaCl content of 1 M. The fractions confirmed to be positive to limulus test with LS-1 kit commercially available from Sei-Ka-
 45 gaku Kogyo Co. in Japan were collected.

The collected fractions were combined and desalted on a column (resin: Sephadex G-25 fine manufactured by Pharmacia in U.S.A.; column size: 2 cm (i. d.) x 25 cm (length); eluting agent: distilled water), and then lyophilized.

- 50 Nucleic acid is of the greatest possibility of being mixed in the lyophilized sample (4.50 mg). Therefore, the UV absorption curve (200 - 400 nm) was prepared, and the absorbance at 60 nm was determined. The nucleic acid content was calculated to be 1% or less on the basis of the above absorbance in view of the fact that the nucleic acid content was 50 µg/ml in the case where the absorbance was 1. In addition, no apparent evidence showing the presence of a protein was observed in SDS electrophoresis. Thus, considering the detection sensibility, the highest content of proteins which may be mixed in the above lyophilized sample was estimated to be 0 - 3 %. Accordingly, the purity of the above lyophilized sample was estimated to be 96 % or more.

The physical properties of the thus prepared B. pertussis LPS were determined in the same manner as described in Reference Example 1. The results were as follows:

Physical properties of B. pertussis LPS

5	Molecular weight:	6,000 \pm 1,000
		9,000 \pm 1,000
		(the data of the two stained bands of the highest stained strength of the plural Coomassie stained bands observed)
	Phosphorus content:	5 per molecular weight of 8,000
	Hexosamine content:	16 \pm 2 per molecular weight of 8,000
	Fatty acid content:	5 per molecular weight of 8,000
10	KDO content:	2 \pm 1 per molecular weight of 8,000

The physical properties of E. coli LPS (0128: B8 manufactured by Difco Co. in U.S.A.) determined in the same manner as described in Reference Example 1 were as follows:

Physical properties of E. coli LPS

15	Molecular weight:	30,000 \pm 5,000
		(the data of the stained band of the highest stained strength of the ladder-like successive Coomassie stained bands)
	Phosphorus content:	12 per molecular weight of 30,000
20	Hexosamine content:	45 \pm 6 per molecular weight of 30,000
	Fatty acid content:	18 per molecular weight of 30,000
	KDO content:	5 \pm 1 per molecular weight of 30,000

Illustrative embodiments of preparations containing LPS according to the present invention will be given in the following examples wherein the LPS content is in terms of E. coli LPS calculated according to limulus test.

Example 1 (tablets)

30	Wheat LPS	0.04 g
	6% HPC lactose	178 g
	Talc stearate	8 g
35	Potato starch	14 g

The above ingredients were mixed and formed into 400 tablets each weighing 0.5 g and containing 0.1 mg of wheat LPS.

Example 2 (solution for internal use)

45	Chlorella LPS	1 mg
	Purified water	100 ml

Example 3 (ointment)

50	Wheat LPS	0.1 g
	Purified lanolin	80 g
55	Yellow petrolatum	ad 1,000 g

Example 4 (injection)

Wheat LPS

0.5 mg

Distilled water for injection ad 1,000 ml

Experiment 1 (quantitative determination of limulus test-positive vegetable LPS)

The quantitative determination of limulus test-positive LPS contained in various plants was carried out using Toxicolor System commercially available from Sei-Kagaku Kogyo Co. in Japan.

1) Distilled water for injection was poured in a flat- or round-bottomed plate having 96 wells in a proportion of 180 $\mu\ell$ per well. Twenty $\mu\ell$ of the test sample (when the sample is a solid, it is dissolved in distilled water for injection) was placed in one of the wells of the plate. Pipetting was effected while stirring with a plate mixer to prepare a ten-fold dilution; hereafter, 20 $\mu\ell$ portions of the respective diluted samples may be collected successively to prepare ten-fold serial dilutions including 100-fold, 1000-fold and so on.

In addition, the degree of dilution may be changed as desired by adjusting the quantitative rate of the distilled water for injection to the sample.

2) As the internal standard, a 100,000-fold dilution of a solution containing 1.5 $\mu\text{g/ml}$ of *E. coli* was prepared and used to confirm that the dilution process and the chromogenic behaviors of the limulus test were normal.

3) Thirty-five $\mu\ell$ of the ten-fold dilution mentioned in the above 1) placed in a well of another plate followed by addition of 35 $\mu\ell$ of LS-1 set of Toxicolor System commercially available from Sei-Kagaku Kogyo Co. in Japan. The mixture was allowed to stand at 37°C for 30 minutes, then 105 $\mu\ell$ of 1 M aqueous acetic acid was added to the mixture to terminate the reaction. The absorbance of this sample solution was determined at a wave length of 415 nm using Plate Reader MTP-100, an absorbance meter for 96 wells manufactured by Corona Denki Co. in Japan. Distilled water was used as the background, and 42 pg/ml of the ET-1 set of the Toxicolor System of Sei-Kagaku Kogyo Co. in Japan was used to prepare a calibration curve. This calibration curve was used as the basis to determine the quantity of the limulus test-positive vegetable LPS of the respective test samples; the absorbance of distilled water was assumed to be 0.

Here, the experiments were carried out again at other dilutions when the value did not come within a range of 10 - 45 pg/ml because the chromogenic strength was confirmed to depend on the content of the limulus test-positive vegetable LPS within said range in the case where the above-mentioned LS-1 set was used.

The quantitative determination of the diluted sample was calculated according to the following equation: (the value determined on the calibration curve) \times (the degree of dilution)

The results of the experiments are shown in Table 2 where solid samples are reported in units of ng/g , and liquid samples, in units of ng/ml .

In the table, the companies and places referred to in the column of samples, are the suppliers and growing districts of the respective samples. Samples without such referring-to were bought at the Nakano-cho branch of the super store Chujitsuya located in Tsukui-gun in Kanagawa, Japan, and their makers could not be identified.

Table 2.

5	Sample (solid)	Content of limulus test-positive LPS (ng)
10	<u>Gymnosperm</u>	
	Pinus spp. (Konan Boeki Co. in Japan)	1 2 5
15	<u>Monocotyledoneae</u>	
	Hard wheat seeds (Chiba Flour Milling Co. Ltd. in Japan)	2 , 2 5 0
20	Hard wheat seeds (m.w.: 5,000 or more) (Chiba Flour Milling Co. Ltd.)	1 , 0 0 0 0 , 0 0 0
25	Hard wheat flour (Chiba Flour Milling Co. Ltd.)	7 , 5 0 0
	Wheat bran (m.w.: 5000 or more) (Chiba Flour Milling Co. Ltd.)	3 0 0
30	Wheat germ (Chiba Flour Milling Co. Ltd.)	1 , 6 0 0
35	Wheat germ (m.w.: 5000 or more) (Chiba Flour Milling Co. Ltd.)	< 1 0 , 0 0 0
	Unpolished rice	1 , 1 0 0
40	Rice flour (m.w.: 5000 or more) (Hinomoto Koku-fun Co. in Japan)	3 1 , 0 0 0 , 0 0 0
	Rice brain	2 9 , 0 0 0
45	Rice brain (m.w.: 5000 or more)	5 0 0 , 0 0 0
	Corn flour (m.w.: 5000 or more) (Taiyo Shiro Co. in Japan)	< 0 . 3

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	Corn grits (m.w.: 5000 or more)	1 2 0
5	(Taiyo Shiro Co. in Japan)	
	Corn (Wako Shokuryo Co. in Japan)	2 0 0
	Gramineae Sasa (Sekimoto Bussan Co. in Japan)	1 5 , 0 0 0
10	Iridaceae Iris (seeds)	3 , 3 0 0
	Garlic (bulb)	7 0
	Asparagus (bud)	4 , 5 0 0
15	Zingiber mloga (flower)	4 1 , 0 0 0
	Colx Lacryma-Jobi L. var. Ma-yuen Stapf	2 , 3 0 0
	(Uchida Wakan-yaku Co. in Japan)	
20	Pinellia ternata (Thunb) Breitenbach	5 , 5 0 0
	(Matsu-ura Yakugyo Co. in Japan)	
	Ophiopogon japonicus Ker-Gawl. var. genuinus	4 , 0 0 0
25	Maxim., O. japonicus Ker-Gawl.	
	(Tochigi Tenkaido Co. in Japan)	
	Turmeric (SB Shokuhin Co. in Japan)	1 9 5 , 0 0 0
30		
	<u>Dicotyledoneae</u>	
35	Soybean (Sanjo Shokuhin Co. in Japan)	1 5 0
	Soybean (m.w.: 5,000 or more)	4 0 0
	(Hokuren Co. in Japan)	
40	"Adzuki" bean (Wako Shokuryo Co. in Japan)	4 5 0
	"Adzuki" bean (m.w.: 5,000 or more)	3 6 , 0 0 0 , 0 0 0
	(Wako Shokuryo Co.)	
45	Broad bean (raw)	7 5 0
	Potato (m.w.: 5,000 or more)	< 0 . 3
	(Hokuren Co.)	
50	Japanese medlar (seed)	8 0 0
55		

	Avocado (seed)	9 5 0
5	Peach (seed)	4 , 5 0 0
	Walnut (seed)	1 , 9 0 0
	Broad bean (seed)	7 5 0
10	Pumpkin (seed)	1 0 , 0 0 0
	Tomato (raw berry)	1 0 , 5 0 0
	"Kaiware daikon" (Japanese radish) (except root)	5 0 , 0 0 0
15	Actinidia polygama Maxim.	4 0 , 0 0 0
	(Marukyu Bussan Co. in Japan)	
	Gynostemma pentaphyllum (Thunb.) Makino	7 3 , 0 0 0
20	(K.K. Sakurai in Japan)	
	Houttuynia cordata Thunb (on wet weight basis)	1 , 2 0 0
	(Medicinal plant garden belonging to Teikyo University in Japan)	
25	White pepper (SB Shokuhin Co.)	2 , 3 0 0
	Capsicum annum L. (Konan Boeki Co. in Japan)	2 , 3 0 0
	Illicium verum Hook. fil. (Konan Boeki Co.)	5 , 5 0 0
30	Nutmeg (Lion Co. in Japan)	2 , 0 0 0
	Sour orange (Uchida Wakanyaku Co.)	8 , 0 0 0
	Kudzu-vine (Tochigi Tenkaido Co.)	3 , 0 0 0
35	Glycyrrhiza glabra L. var. glandulifera Regal	1 8 , 0 0 0
	et Herder (Uchida Wakanyaku Co.)	
40	Carrot (Uchida Wakanyaku Co.)	4 5 , 0 0 0
	Seseli libanotis Koch var. daucifolia DC.	5 0 , 0 0 0
	(Tochigi Tenkaido Co.)	
45	Sinomenium acutum Rehd. et Wils	6 0 0 , 0 0 0
	(Tochigi Tenkaido Co.)	
	Uncaria rhynchophylla Miq., Ourouparia	7 , 0 0 0
50	rhynchophylla Matsum. (Uchida Wakanyaku Co.)	

	Hachimi-jiwogan (Kanebo Yakuhin in Japan)	1 7 , 0 0 0
	Shosaikoto (Tsumura in Japan)	1 3 , 0 0 0
5	Goreito (Tsumura in Japan)	1 2 , 0 0 0
	Choreito (Tsumura in Japan)	1 4 , 0 0 0
	Juzendaihoto (Tsumura in Japan)	8 , 0 0 0
10	Hachimi-jiwogan (Tsumura in Japan)	8 , 0 0 0
	Royal Jelly (Pekin Royal Jelly)	1 , 0 0 0
15	Honey (Kato Bihoen Honpo Co. in Japan)	8 0 0

Pteridophyta

20	Horse tail (on wet weight basis)	7 0 0
	(Medicinal plant garden belonging to Teikyo University in Japan)	
	Royal fern (Sekimoto Bussan Co. in Japan)	1 0 , 0 0 0

25

Algae

	Undaria pinnatifida Suringar	1 1 , 0 0 0
30	(Sanriku district in Japan)	
	Bud of Undaria pinnatifida Suringar	2 0 0 , 0 0 0
	(Moriya Kenko Shokuhin Co. in Japan)	
35	Hijikia fusiformis (raw)	8 5 , 0 0 0
	Bud of Hijikia fusiformis	1 0 5 , 0 0 0
	(Sho-zen Hon-ten in Japan)	
40	Kelb (Yamato Takahasi Co. in Japan)	2 3 5 , 0 0 0
	Asakusa laver (dried raw laver)	1 3 0 , 0 0 0
	Chlorella	
45	(Healstar Japan YS Co. in Japan)	1 , 9 0 0 , 0 0 0
	(Mannan foods YS Co. in Japan)	1 , 0 0 0 , 0 0 0

50

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Fungi

	Lentinus edodes Sing., Cortinellus shiitake	1 6 , 0 0 0
5	P. Henn. (Shimonita in Shizuoka, Japan)	
	Winter mushroom (Nakano City in Nagano, Japan)	2 0 , 0 0 0
10	Lyophyllum shimeji (Seta-gun, Miyagi-machi in Gunma, Japan)	4 0 , 0 0 0
	Polyporales Grifola (Ohtone in Japan)	2 0 5 , 0 0 0
15	Awabitake (Hanyu in Japan)	8 , 0 0 0
	Mushroom	2 0 , 0 0 0
	Jew's ear	7 5 , 0 0 0
20	Pholiota nameko	2 1 , 0 0 0
	Ebios	2 5 0 , 0 0 0
	(Brewer's yeast manufactured by Asahi Beer Co. in Japan)	
25	Cordyceps sinensis Sacc., Cordyceps sobolifera	2 4 0 , 0 0 0

30

Limulus test-positive

Sample (Liquid)LPS (ng)

35

Beer

40

Kirin's	FINE PILSNER	1 , 1 5 0
	LAGER BEER	1 , 2 5 0
	HEARTLAND	1 , 5 5 0
	FINE DRAFT	1 , 4 0 0
Asahi's	SUPER YEAST	6 0 0

45

Wine

Suntory's Ste. Neige (white)	1 3
(red)	2 4
50 Cidre (apple)	9 0 0

55

Sake (Japanese liquor)

	Ozeki, first grade (Ozeki Shuzo Co.)	2 . 4
5	Kizakura, second grade (Kizakura Shuzo Co.)	1 . 7
	Taikan Ginjo, second grade (Gyokusendo Shuzo Co.)	2 . 1

10 Sake from unpolished rice

	Hibi Ikkon (Ozeki Shuzo Co.)	1 2
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15 Herb liquor

	Totoshu DELCUP (Totoshu Honpo Co. in Japan)	1 . 2
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20 Shochu (Japanese low-class distilled spirits)

	TAKARA SHOCHU (Takara Shuzo Co. in Japan)	< 2 . 0
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25 Others

	KYOLEOPIN (Wakunaga Seiyaku Co. in Japan)	6 0 0
	Garlic extracts (Wakunaga Seiyaku Co. in Japan)	3 5 0
30	Gross Q (Chlorella Kogyo Co. in Japan)	6 , 0 0 0
	Ohmugi Kenko Mekkoru (Ichiwa in Korea)	2 , 0 0 0
	Sacron Herb Solution (Ezai in Japan)	1 , 0 0 0
35	Extracts of dishcloth gourd	7 0 0
	Bio-arugen (Chlorella Kogyo Co. in Japan)	4 0 0
	Pan-siron Naifukueki (Rhoto Seiyaku Co. in Japan)	2 0 0
40	Yunkeru Fantie (Sato Seiyaku Co. in Japan)	5 0
	Korihogusu (Kobayashi Seiyaku Co. in Japan)	3 0
	Today (Sankyo in Japan)	2 0
45	Mio D Kowa 100 (Kowa in Japan)	1 0
	Ri-gein (Sankyo in Japan)	9

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	Roburen 50 (Dai-ichi Seiyaku in Japan)	7
	Sorumack (Taiho Seiyaku in Japan)	6
5	Ro-jerry Gold (Chugai Seiyaku in Japan)	5
	Pas-bitan 30 (Tokiwa Seiyaku in Japan)	5
	Thio-bita (Taiho Seiyaku in Japan)	5 or less
10	Ripobitan (Taiho Seiyaku in Japan)	5 or less
	Aspara Gold (Tanabe Seiyaku in Japan)	5 or less

15 Experiment 2 (choice of LPS whose macrophage activation ED_{50} is 0.4 - 100 ng/ml of culture solution in terms of its limulus test-positive LPS content)

Two hundred $\mu\ell$ (2×10^5 cells) / well of macrophage peritoneal cells of mice (each group comprised three mice of 9 weeks old and having an average weight of 29 g) were placed in a flat-bottomed plate having 96 wells, and 10 $\mu\ell$ of recombinant IFN- γ (100 units/ml) as a primer was placed in the respective wells. Separately, extracts prepared by extraction of various LPS sources with hot water at 65°C for 5 hrs. were diluted to various degrees, and then were administered as triggers to the cells three hours after the administration of the primer in a proportion of 10 $\mu\ell$ /well. After two hour culture, the culture solutions were subjected to centrifugation at 3,000 G for 20 min. A pipette was used to collect 130 $\mu\ell$ of the supernatant from the respective wells, and the TNF activity was determined on the basis of the toxicity to L929 cells. The limulus test-positive LPS content was determined using Toxicolor System[®] commercially available from Sei-Kagaku Kogyo Co. in Japan.

The values were plotted to prepare a graph wherein the quantity of the TNF produced (units / ml of culture solution) was plotted along the axis of ordinate, whereas the corresponding limulus test-positive LPS content (ng / ml of culture solution) was plotted along the axis of abscissa (on a logarithmic scale), and a sigmoid curve was prepared on the basis of the plotted points. The macrophage activation ability was estimated to be 0 % in the case where it corresponded to the quantity of TNF produced by macrophage with no trigger added thereto, and 100 % was assigned to the macrophage activation ability which provided the maximal constant of TNF. The limulus test-positive LPS content supposed to provide 50% of macrophage activation ability was determined with reference to the curve.

35 Table 3 given below shows the data of the LPS sources satisfying the above-mentioned correlation between the macrophage activation ability and the limulus test-positive LPS content. In the table, "TNF", "activation ability" and "LPS" represents the quantity of the TNF produced (units / ml of culture solution), macrophage activation ability (%) and limulus test-positive LPS content (ng / ml of culture solution), respectively. Here, the quantity of the TNF produced in the case where no trigger was added was 0.75 units / ml, so the macrophage activation ability was determined to be 0 % in the case where the quantity of the TNF produced was 0.75 units / ml or less. The macrophage activation ability (%) was calculated according to the following equation:

$$\frac{\text{Quantity of TNF produced} - 0.75}{\text{Maximal constant of TNF} - 0.75} \times 100$$

Table 3

LPS sources	TNF	Activation ability	LPS
Turmeric	0.75	0	0
	3.9	9	0.6
	36.3	100	60
	36.3	100	>1000
Sinomenium acutum Rehd. et Wils.	0.75	0	0
	40.7	100	4
	36.5	90	400
	40.7	100	>1000
Kelp (Laminaria japonica)	0.75	0	0
	1.3	4	0.8
	13.0	100	80
	13.0	100	>1000
Asakusa laver (Porphyra tenera)	0.75	0	0
	1.0	2	0.3
	12.8	100	30
	12.8	100	>1000
Extracts of Undaria pinnatifida Suringar (bud)	0.75	0	0
	1.3	4	0.2
	15.5	100	20
	15.5	100	>1000
Hijikia fusiformis (bud)	0.75	0	0
	5.7	8	0.7
	62.7	100	70
	62.7	100	>1000
Ebios	0.75	0	0

	0.6	0	0.7
	30.6	100	70
5	30.6	100	>1000
	Cordyceps sinensis Sacc., Cordyceps sobolifera	0.75	0
		2.0	0.4
10		30.3	40
		30.3	>1000
	Undaria pinnatifida Suringar (bud)	0.75	0
15		0.9	0.4
		22.7	40
20		22.7	>1000
	Chlorella	0.75	0
		39.2	9.6
25		35.0	960
	E. coli LPS	0.75	0
		3.6	2
30		10.2	20
		11.4	200
35		10.9	2000
	Wheat LPS	0.75	0
		0.7	2
40		10.1	21
		10.2	210
45		8.5	2100
	B. pertussis	0.75	0
		0.7	11
50		3.3	110
		5.4	1100

Lipid A	0.75	0	0
	4.7	37	2
	9.4	80	24
	11.1	96	240
	11.5	100	2400

The results given in Table 3 are shown in Figs. 5 - 8.

In Figs. 5 - 8, the axis of ordinate represents the macrophage activation activity (%), whereas the axis of abscissa (on a logarithmic scale) represents the limulus test-positive LPS content (ng / ml of culture solution).

In Fig. 5, ○, ●, □, and ■ show the data of Turmeric, *Sinomemium acutum* Rehd. et Wils., Kelp and asakusa laver, respectively.

In Fig. 6, ○, ● and □ show the data of extracts of *Undaria pinnatifida* Suringar (bud), *Hijikia fusiformis* (bud) and Ebios, respectively.

In Fig. 7, ○, ● and □ show the data of *Cordyceps sinensis* Sacc., *Cordyceps, sobolifera*, *Undaria pinnatifida* Suringar (bud) and *Chlorella*, respectively.

In Fig. 8, ○, ●, □, and ■ show the data of *E. coli* LPS, wheat LPS, *B. pertussis* LPS and lipid A, respectively.

Experiment 3

(determination of effect to lower cholesterol level)

To female WHHL rats having an average body weight of 4 kg, there was given only drinking water (control group) or drinking water containing 1 mg (900 μg / g in terms of limulus test-positive LPS) / ml of Powder A-a₂ prepared as described in Reference Example 1 at will over a period of 45 days. The quantity of the water taken was about 500 ml per day in the respective groups; no significant difference was found. Further, the body weight of the animals changed little during the experiment.

The serum cholesterol level was determined using V cholestase (Nissui Co. in Japan). The results are shown in Table 3 given below as an average of the three. In the table, "the start" means the start of the medication.

Table 3 (Serum cholesterol level: mg/ml)

Determined	Control group	Test group
7 days before the start	6 7 3 . 2	4 1 3 . 6
1 day before the start	6 7 3 . 2	3 7 5 . 1
2 days after the start	5 9 6 . 2	3 0 8 . 0
9 days after the start	7 0 1 . 8	3 0 8 . 0
16 days after the start	6 0 6 . 1	2 5 0 . 0
23 days after the start	5 7 7 . 5	2 3 1 . 0
30 days after the start	5 8 7 . 4	1 8 2 . 6
37 days after the start	6 1 6 . 0	2 6 9 . 5
44 days after the start	6 3 4 . 7	2 2 1 . 2
50 days after the start	5 6 7 . 6	2 1 1 . 2
57 days after the start	6 3 7 . 4	2 0 2 . 4

Fig. 9 is a graph showing the results given in Table 3. In Fig. 9, ● represents the data of the control group to which only drinking water was given, while ○ represents the data of the test group to which the cholesterol lowering agent of the present invention was given.

Fig. 9 demonstrates the excellent and lasting effect of the cholesterol-lowering agents and veterinary cholesterol-lowering agents of the present invention to lower cholesterol level.

Dose, interval and toxicity

The dose and the interval of the cholesterol-lowering agents and veterinary cholesterol-lowering agents of the present invention are of course determined by the doctor or veterinarian in charge individually in view of the age, conditions, etc. of the patient. However, it may be said that 1 μ g - 100 mg (oral administration), 10 ng - 1 mg (intravenous administration) and 100 ng - 1 mg (percutaneous administration) are standard single dose per day to adults (body weight 60 kg). For veterinary use, about one sixtieth of the above quantities may be given per 1 kg of body weight of large-sized animals such as cattle, horses or the like. About twice as much as the dose to large-sized animals may be given per 1 kg of body weight of medium- or small-sized animals such as pigs, dogs, cats or the like. Fowls or the like may receive twice as much as the dose to medium- or small-sized animals.

Wheat LPS (Reference Example 1), Chlorella LPS (Reference Example 2), E. coli LPS (0128:B8 manufactured by Difco Co. in U.S.A.) and B. pertussis LPS had the following LD₅₀ (an average of the data on two male BALB/C mice weighing 45 kg on average).

LPS	LD ₅₀ / kg (mg)	
	i.v.	i.c.
Wheat LPS	3 . 2	1 6
E. coli LPS	3 . 4	1 6
B. pertussis LPS	1 1	3 2

15 **Claims**

1. A cholesterol-lowering composition comprising an effective amount of at least one member of LPS whose macrophage activation ED₅₀ is 0.4 - 100 ng/ml of culture solution in terms of its limulus test-positive LPS content observed on a sigmoid curve prepared by determining the ability of the LPS to activate the TNF productivity of macrophage cultured *in vitro*, and plotting the macrophage activation ability (%) along the axis of ordinate wherein the ability is estimated to be 0 % in the case where it corresponds to the quantity of TNF produced by macrophage with no LPS added thereto, and 100 % is assigned to the macrophage activation ability which provides the maximal and constant quantity of TNF produced by the macrophage and plotting the limulus test-positive LPS content of the LPS along the axis of abscissa on a logarithmic scale, in admixture with a pharmaceutically or veterinarily acceptable carrier, such that when administered to an animal, high cholesterol level of said animal is prevented or cured.
2. The cholesterol-lowering composition of Claim 1, wherein the LPS is selected from the group consisting of vegetable LPS, bacterial LPS and lipid A.
3. The cholesterol-lowering composition of Claim 2, wherein the vegetable LPS is wheat LPS.
4. The cholesterol-lowering composition of Claim 3, wherein the wheat LPS is the one having the following physical properties.
 - Molecular weight: 8,000 ± 1,000 (on SDS electrophoresis)
 - Phosphorus content: not less than 1 per molecular weight of 8,000
 - Hexosamine content: 6 ± 2 per molecular weight of 8,000
 - Fatty acid content: 6 ± 2 per molecular weight of 8,000
 - KDO content: 5 ± 1 per molecular weight of 8,000
5. A composition for treating high cholesterol level of an animal comprising an effective amount of the composition of claim 1 to prevent or cure the high cholesterol level of said animal.

FIG. 1

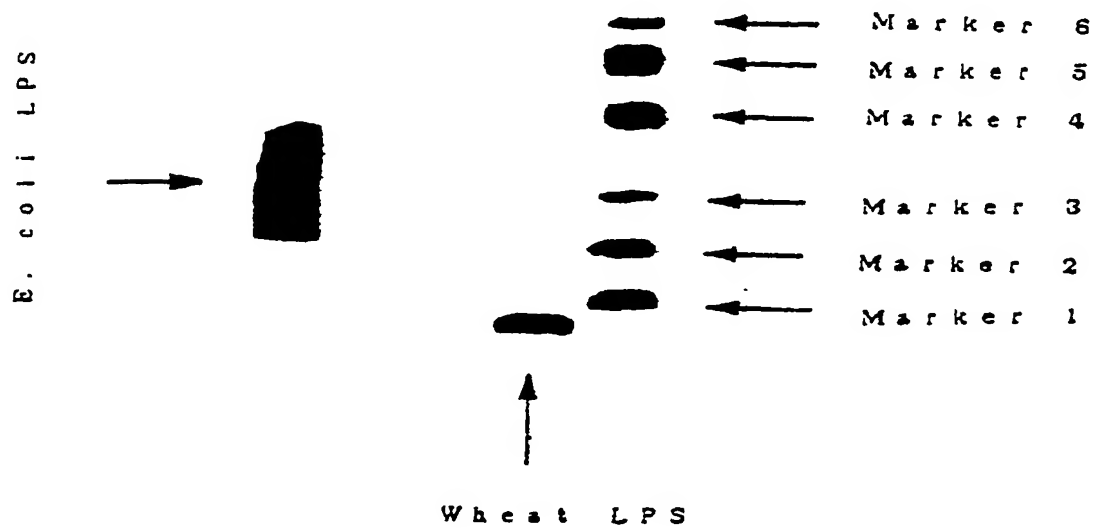


FIG. 2

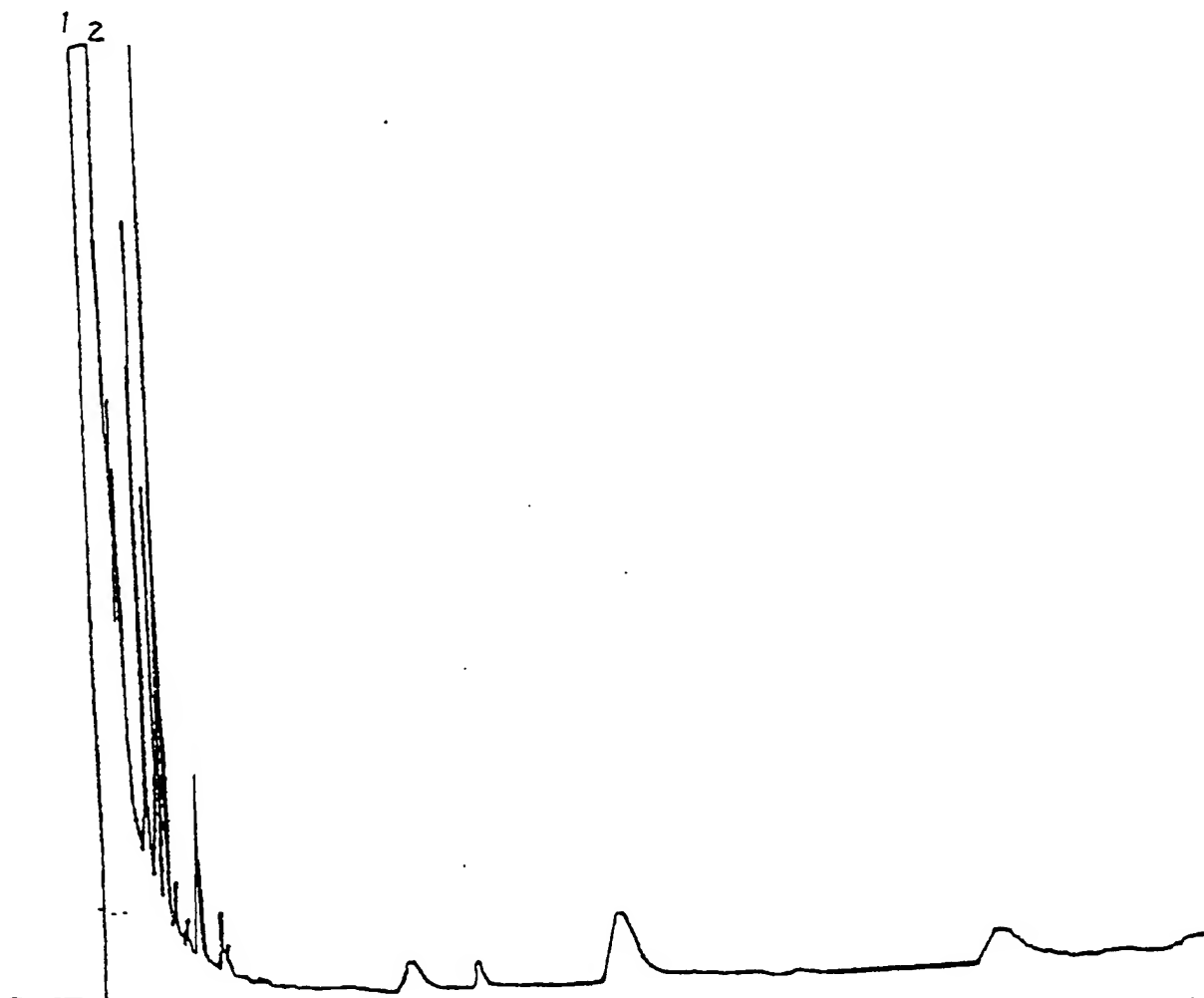


FIG. 3

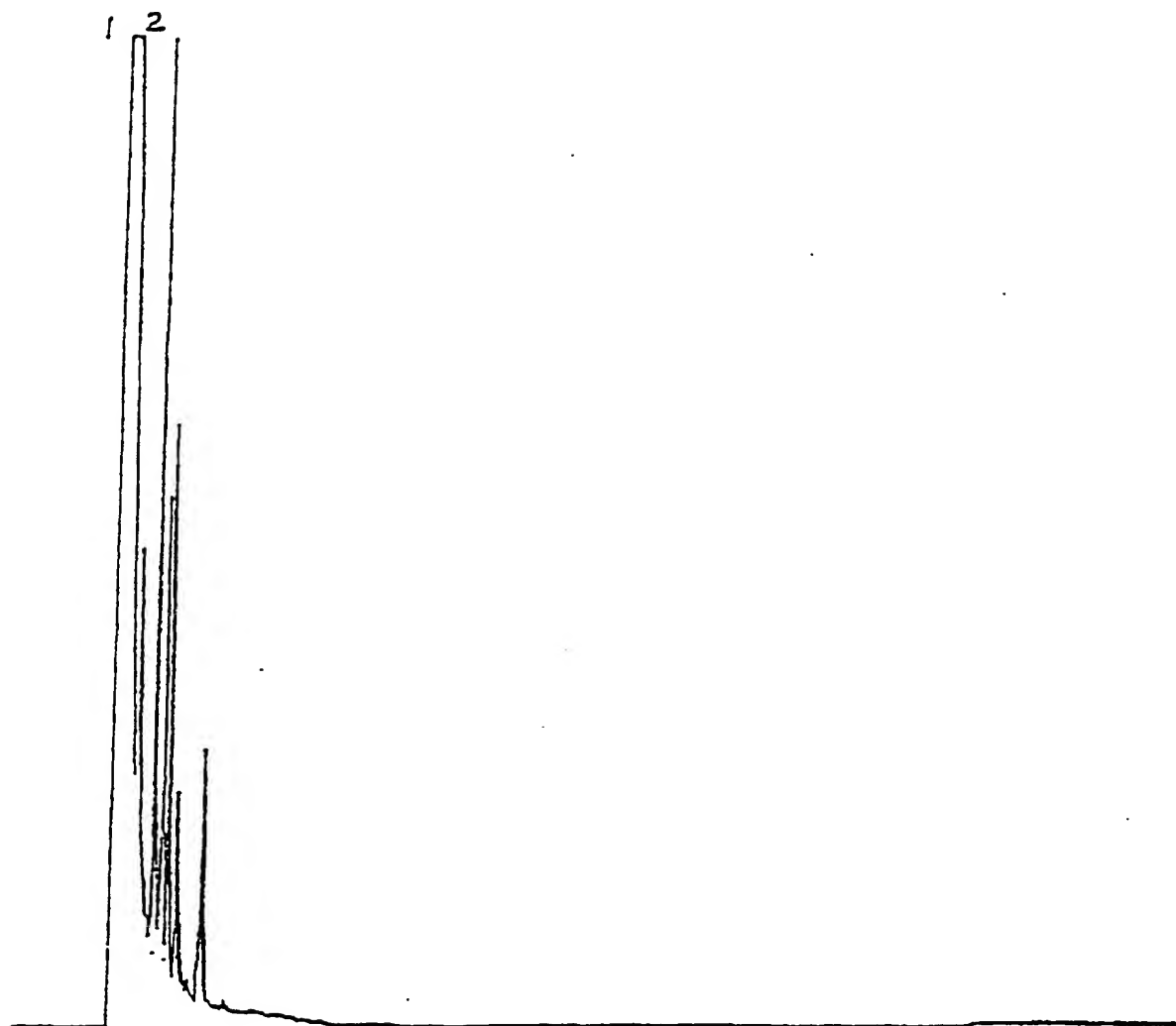


FIG. 4

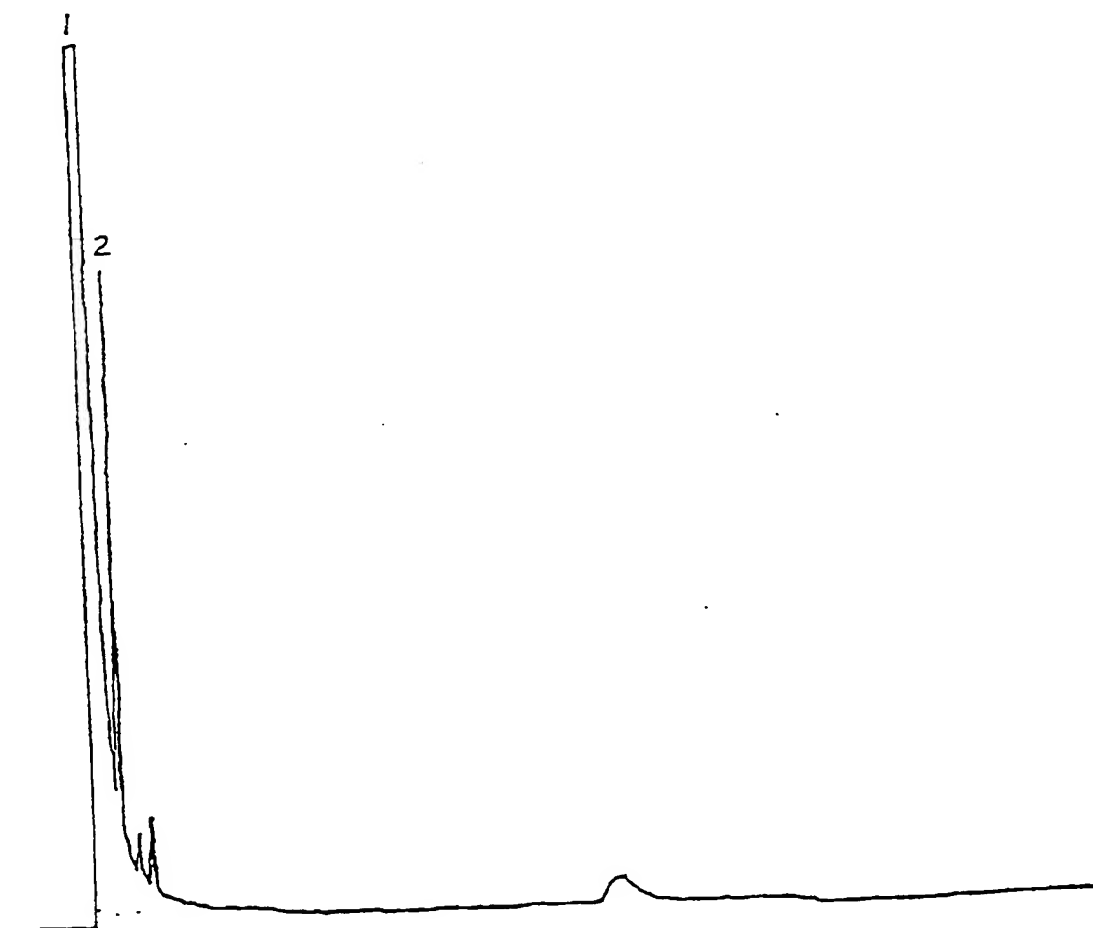


FIG. 5

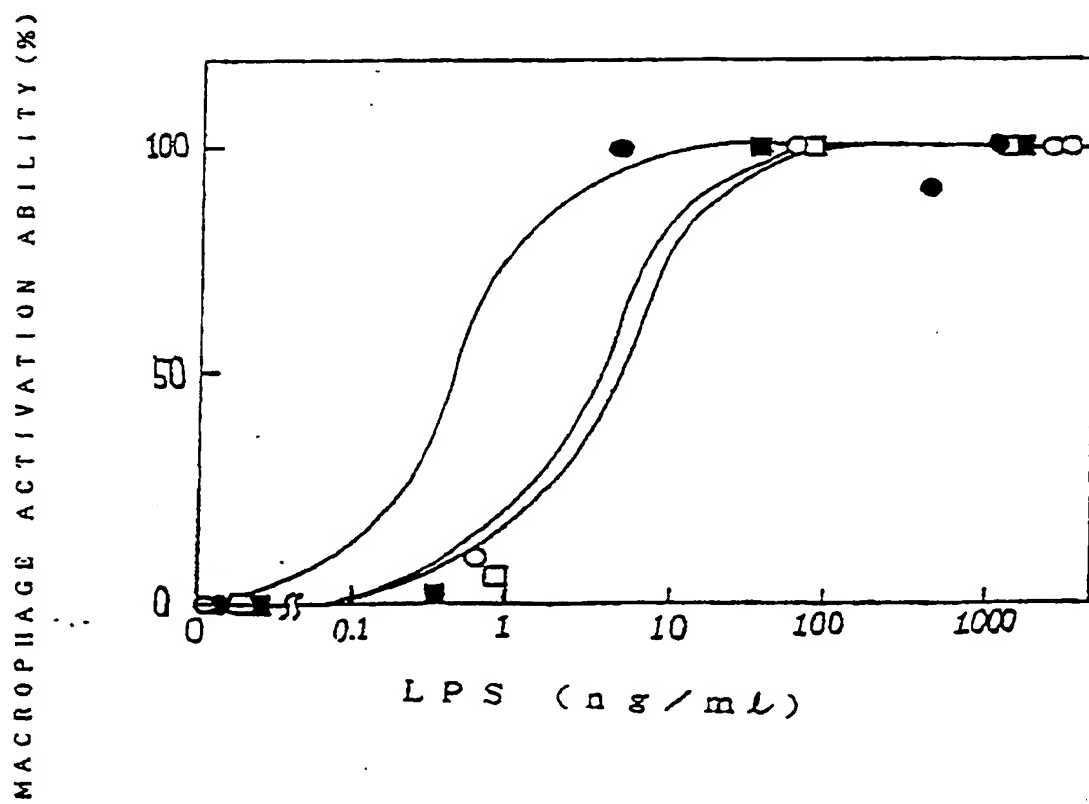


FIG. 6

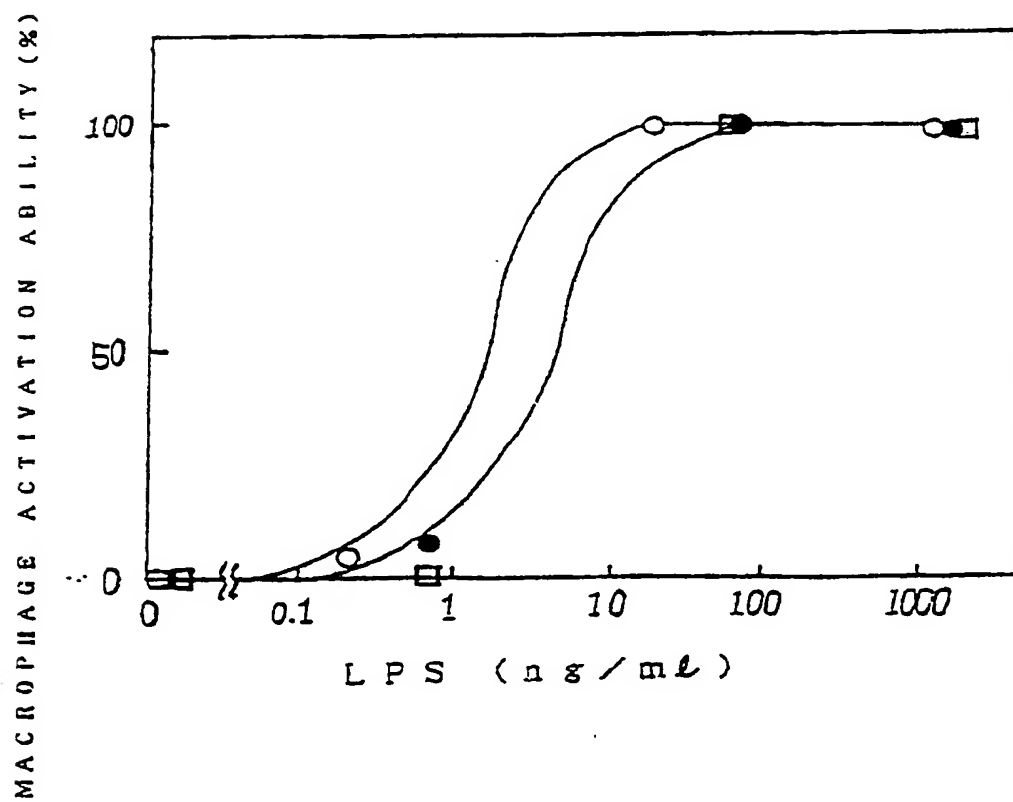


FIG. 7

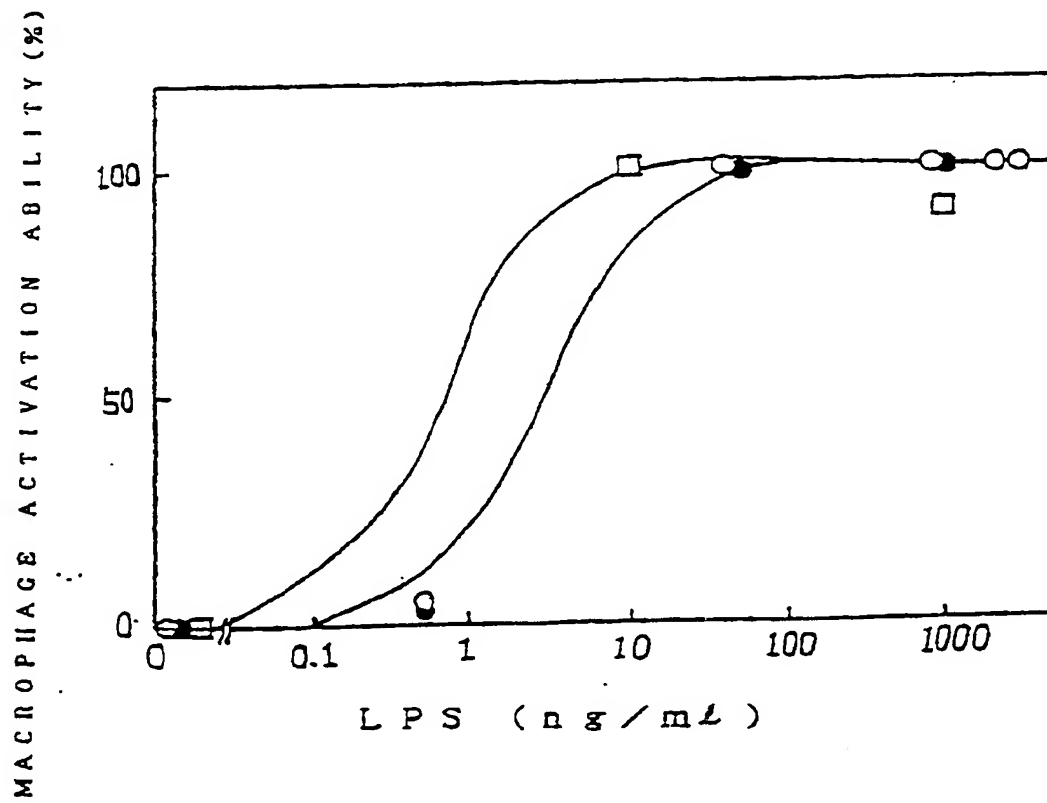


FIG. 8

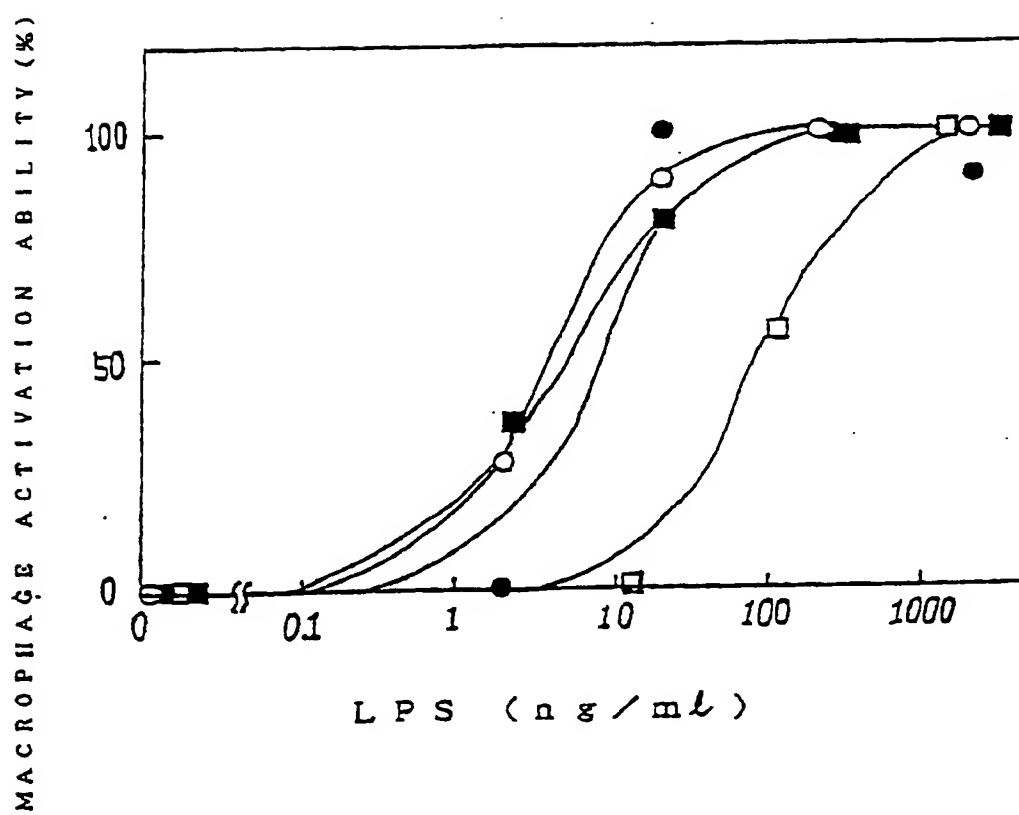


FIG. 9

